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Bibliographie

Chailey, M. (1980) Notes sur les Afars de la région de Tadjoura. Soc. d'Etude de l'Afrique Orientale.

Hervé, M. et Dancet, M. (1978) Enquête sanitaire et socioéconomique. Projet Nomadep, Ethiopie.

Kelecha, M. (1977) A Glossary of Ethiopian Plant Name. 2nd edn. revised. Addis Abeba, Ethiopia.

Kokwaro (1976) Medicinal Plants of East Africa. East African Literature Bureau, Nairobi. Lemordant, D. (1960) Les Plantes Éthiopiennes. Central Printing Press, Addis Abeba,

Lemordant, D. (1971) Contribution à l'ethnobotanique éthiopienne. Journal d'Agriculture Tropicale et de Botanique Appliquée 18 (1 à 6), 1-35 et 142-179.

Lemordant, D. (1981) Plantes toxiques d'Ethiopie. Fitoterapia. LII (3), 99-127.

Touzeau, J. (1973) Les arbres fourragers de la zone sahélienne de l'Afrique. Thése Doct. Vétérinaire, Toulouse.

Short Communication

PHYTOCHEMICAL INVESTIGATION OF TABERNAEMONTANA CRASSA*

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Summary

From the stembark of Tabernaemontana crassa the alkaloid ibogaine was isolated as the major component. Ibogaine showed activity against the grampositive Bacillus subtilis. Conopharyngine was identified as one of the minor compounds.

Introduction

Tabernaemontana crassa Benth. (Apocynaceae) occurs widely in tropical Africa from Sierra Leone to Uganda in the north and to Angola and Tanzania in the south. It is a bush or small tree which grows particularly in sparse forest or in secondary vegetation from the coast up to an altitude of 2000 m. The species has many synonyms, the most important of which are T. durissima Stapf, T. jollyana Pierre ex Stapf, Conopharyngia crassa (Benth.) Stapf, C. durissima Stapf and Gabunia odoratissima Stapf (van Beek et al., 1984a). Various parts of this species are widely used in traditional medicine for several different ailments (van Beek et al., 1984a). In a recent antimicrobial screening of 19 Tabernaemontana species (van Beek et al., 1984b) the ethanolic stembark extract of T. crassa showed significant activity against gram-positive bacteria and weak activity against gramnegative bacteria. This fact led to the present study, the results of which are described below. Previous phytochemical investigations of the stembark

^{*}Part 12 in the series: Pharmacognostical studies of Tabernaemontana species. For part 11 see van Beek et al. (1985).

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nave yielded the alkaloids O-acetyl-polyneuridine, anhydro-vobasindiol, akuammiline, conopharyngine, 19-hydroxy-conopharyngine, 3-oxo-conopharyngine, conopharyngine-hydroxy-indolenine, coronaridine, 3-oxo-coronaridine, 5-oxo-coronaridine, 3-oxo-coronaridine-hydroxy-indolenine, heyneanine, 3-oxo-heyneanine, isovoacangine, voacristine and crassanine (van Beek et al., 1984a).

Materials and methods

The plant material was collected in Cameroon, 3 km from Rocher du Loup by H. Beentje in December 1980. The plant material was identified by Dr. A.J.M. Leeuwenberg and a voucher specimen (H. Beentje 1548) has been deposited at the Laboratory for Plant Systematics, Wageningen, The Netherlands. The 'H-NMR spectrum was recorded at 300 MHz in CDCl₃ (Bruker WM 300). The MS was recorded at 70 eV using a direct inlet system (AEI MS 20). The following TLC systems were used in combination with silica gel plates: (A) cyclohexane/CHCl₃/Et₂NH (6:3:1); (B) EtOAc/isoPrOH (9:1); (C) toluene/EtOH satd. with NH₃ (19:1) (prior to development the plates were left standing in an atmosphere of NH₃ for 20 min). (D) CHCl₃/MeOH (9:1); (E) EtOAc/isoPrOH/26% NH₄OH (45:4:1). After development the TLC plates were sprayed with iodoplatinate reagent, 1% Ce(SO₄)₂ in 10% H₂SO₄ or with 0.2 M FeCl₃ in 35% HClO₄ followed by heating with hot air.

Isolation procedure: The ground stem bark (800 g) was extracted for 15 h with 96% EtOH in a Soxhlet apparatus working under a pressure of 0.2 atm. After cooling at -16° C a precipitate was formed, which was collected (steroids and triterpenes). The remaining solution was evaporated in vacuo to dryness (10 g). Five grams of the extract was partitioned between CHCl₃ and 2% HOAc. The aqueous layer was collected and adjusted to pH 10 with NH₄OH and extracted twice with CHCl₃. The CHCl₃ layer was collected, dried (Na₂SO₄) and evaporated in vacuo (yield 0.97 g tertiary alkaloids, 0.24%). Part (100 mg) of this fraction was separated by means of preparattive TLC with systems A and E. The major amorphous product (12 mg) was identified as ibogaine (1) and showed the following physical data: R_f values and chromogenic reactions (see van Beek et al., 1984c); UV λ_{max} (MeOH) 227 and 297 nm; MS (175°C) m/z (rel. int.) 310 (M⁺, 70), 309 (27), 295 (12), 225 (39), 149 (46), 136 (100), 135 (78), 122 (48); ¹H-NMR: 7.52 (bs. NH), 7.14 (d, J = 8.8 Hz, H12), 6.93 (d, J = 2.5 Hz, H9), 6.76(dd, J = 8.8 and 2.5 Hz, H11), 3.85 (s, OMe), 3.42-3.38 (m, H5a and H5b),3.18-3.10 (m, H6a), 3.07 (ddd, J = 9.5, 2.1 and 2.1 Hz, H3a), 2.97 (ddd, J = 9.5, 3 and 3 Hz, H3b), 2.90 (ddd, J = 11.6, 4.2 and 1.5 Hz, H16), 2.85 (bs, H21), 2.65-2.57 (m, H6b), 2.04 (dddd, J = 13.0, 11.6, 2.6 and 2.6 Hz, H17a), 1.84 (bs, H14), 1.81 (m, H15a), 1.65 (m, H17b), 1.63-1.42 (m, H19a, H19b and H20), 1.21 (m, H15b), 0.89 (J = 7.2 Hz, H18).

$$R_1$$
 R_2
 N
 R_3

Fig. 1. 1: $R_1 = OMe$; $R_2 = R_3 = H$. 2: $R_1 = R_2 = OMe$; $R_3 = CO_3Me$.

Antimicrobially inactive minor alkaloid: This alkaloid was identified by means of coTLC as conopharyngine (2). R_f -values and chromogenic reactions see van Beek et al. (1984c).

Antimicrobially active minor alkaloid: R_f -value in system A 0.43, C 0.58, D 0.09. Colour with the FeCl₃ spray reagent: blue before and after heating.

Results and discussion

A preliminary TLC investigation of the crude tertiary alkaloid fraction showed that one major alkaloid, two minor alkaloids and many trace alkaloids were present. Using the agar diffusion method (Verpoorte et al., 1982) as a bioassay it was found that the major alkaloid and one of the minor alkaloids showed activity against Bacillus subtilis. The activity of the major alkaloid was in this bioassay comparable with the activity of 3-R/S-hydroxy-isovoacangine which has a MIC value of 50 µg/ml against B. subtilis. The major alkaloid was isolated by means of preparative TLC and identified by means of its spectral data (UV, MS and ¹H-NMR) and coTLC as ibogaine (1). The antimicrobially active minor alkaloid was present in too low a concentration to allow its isolation. Its R_f -values and chromogenic reactions suggested that it was an alkaloid of the dimeric voacamine type lacking the carbomethoxy group at the 16-position of the iboga half (van Beek et al., 1984c). The other minor alkaloid was identified by means of coTLC and its chromogenic reactions as conopharyngine (2). We do not exclude the possibility that some of the trace alkaloids may also contribute to the overall antimicrobial activity of this species. This could not be checked because of the low sensitivity of the biogram method. Ibogaine (1) has not previously been isolated from the stembark or any other part from this species.

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- van Beek, T.A., Verpoorte, R. and Baerheim Svendsen, A. (1985) Antimicrobially active alkaloids from Tabernaemontana chippii. Journal of Natural Products 48, 400-423.
- van Beek, T.A., Verpoorte, R., Baerheim Svendsen, A., Leeuwenberg, A.J.M. and Bisset N.G. (1984a) Tabernaemontana L. (Apocynaceae): A review of its taxonomy, phytochemistry, ethnobotany and pharmacology. Journal of Ethnopharmacology 10, 1—156.
- van Beek, T.A., Deelder, A.M., Verpoorte, R. and Baerheim Svendsen, A. (1984b) Antimicrobial, antiamoebic and antiviral screening of some *Tabernaemontana* species. Planta Medica 50, 180-185.
- van Beek, T.A., Verpoorte, R. and Baerheim Svendsen, A. (1984c) Identification of Tabernaemontana alkaloids by means of thin-layer-chromatography and chromogenic reactions. Journal of Chromatography 298, 289-307.
- Verpoorte, R., Ruigrok, C.L.M. and Baerheim Svendsen, A. (1982) Medicinal plants of Surinam II, antimicrobially active alkaloids from Aspidosperma marcgravianum. Planta Medica 46, 149-152.

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Book Review

Alkaloids: Chemical and Biological Perspectives, Vol. 2, S.W. Pelletier (Ed.) John Wiley & Sons, New York, 1984, xi + 490 pp.

A worthy continuation of the numerous contributions to our knowledge of alkaloids made by Dr. S. William Pelletier of the Institute of Natural Products Research and the University of Georgia, this volume goes a step ahead and gathers together unpublished as well as summaries of hard-to-locate published material not only of purely chemical and biological interest but data indicative of the editor's breadth of interest. It succeeds in convincing the specialist that this field of research must be viewed from the interdisciplinary viewpoint.

The physiological effects and social importance of some of the 5000 known alkaloids are broadly discussed. Yet with this broad approach, the book succeeds admirably in offering a vast amount of highly technical chemical and biological information, a good portion of it new.

The volume treats five aspects of alkaloid study, the work of 12 authors—all highly qualified specialists. Chapter 1 reviews the applications of single-cell X-ray diffraction in alkaloid chemistry. Chapter 2 surveys the imidazole alkaloids. Chapter 3 presents a comprehensive review of quinolizidine alkaloids of the Leguminosae. In Chapter 4, a broad summary of the chemistry and pharmacology of maytansinoid alkaloids is offered. Chapter 5 concerns ¹³C and proton NMR shift assignments and physical contents of C₁₉-diterpenoid alkaloids.

The two indices, one for subjects and one for organisms, materially increase the availability of the vast amount of material presented.

This is a book which the ethnopharmacologically oriented phytochemist, pharmaceutical scientist or botanist should have at hand. It includes much information so efficiently organized that utility is greatly enhanced and modern concepts are fully summarized.

Richard Evans Schultes