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arrangement and hydrogen transfer from the dihydropyridine to the acrylic ester function yielding the hemiketal X with loss of 3-ethylpyridine as indicated. Elimination of methanol and further rearrangement then give the carbazole VIII. In support of the latter process 1-methyl-2-methoxycarbazole (IX) could be detected and characterized as a minor product of the reaction.



More direct evidence for the formation of I was obtained by the capture in 50% yield of the racemic salt XI when catharanthine was heated in methanol at 140° for 2 hr. The pyridinium salt had λ_{max} (EtOH) 219, 269, 283 (sh), 291 nm; $[\alpha]_{300-600} 0^{\circ}$ (EtOH); nmr⁸ $(D_2O) \tau$ 1.7-3.3, 8 H, m (Ar-H); 5.30 and 6.66, 4 H, 2t ($-CH_2CH_2-$); 6.12, 1 H, q, J = 7.5 Hz (CH- $(CH_3)CO_2CH_3$; 6.33, 3 H, s (CO_2CH_3) ; 7.55, 2 H, $q, J = 7.5 Hz (CH_2CH_3); 8.62, 3 H, d, J = 7.5 Hz (CH (CH_3)CO_2CH_3$; 9.18, 3 H, t, J = 7.5 Hz (CH_2CH_3) .

In contrast to the intramolecular formation of the carbazole from the dihydropyridineacrylic ester I in the aprotic solvent xylene, the availability of solvent protons in the latter case appears to divert the collapse of this intermediate in methanol via an ionic mechanism to the pyridinium salt XI.⁹ This salt is stable in methanol at 175° but on pyrolysis at this temperature affords the carbazole VIII, presumably via elimination of ethylpyridine and cyclization of the resulting vinyl ester XII. The generation of I in methanol solution could also be rationalized by an ionic mechanism⁴ which recalls the formation of the betaine XIII from akuammicine (XIV).¹⁰ Since the species I and XI could be reached in vivo from stemmadenine,⁴ tabersonine (II), and catharanthine (III), it will be of interest to test these three alkaloids as biochemical precursors for V and VI and also to consider the system I \rightleftharpoons XI as a labile but isolable biosynthetic intermediate for the Aspidosperma and Iboga alkaloids. These experiments also

(9) When the reaction is carried out in CH₃OD solution, the nmr spectrum of the salt no longer shows a signal at τ 6.12 (CD(CH₃)CO₂-CH_a) and the doublet at τ 8.62 (CH(CH_a)CO₂CH_a) is replaced by a singlet (3 H). These observations are in accord with the mechanism $I \rightarrow$ XI.

(10) P. N. Edwards and G. F. Smith, J. Chem. Soc., 1458 (1961).

provide a mechanistic rationale for the previous vitro transformations⁴ which have been found to k particularly sensitive to the temperature employed the reaction⁷ and where merging electrocyclic and ionic mechanisms may be operating.

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Biogenesis of Strychnos, Aspidosperma, and Iboga Alkaloids. The Structure and Reactions of Preakuammicine Scott & Qureshi

Sir:

In the course of biosynthetic and biogenetic investig tion of the indole alkaloid family, models have been devised 1-3 to simulate the Corynanthe \rightarrow Strychnos \rightarrow Aspidosperma \rightarrow Iboga transformations. However the information revealed by these and related feeding experiments^{4,5} still leaves unanswered both the nature of the intermediates between geissoschizine (I) (Corp. nanthe), akuammicine (II) (Strychnos), and stemmade nine (III) ("Corynanthe-Strychnos") and the mecha nisms connecting them. It has been suggested[®] that key member of this family situated at the main branch ing point of the pathway would be a prototype (IM) of the Strychnos alkaloids retaining all ten of the original geraniol carbon atoms which could suffer irreversible loss of a single carbon function to the "C₉" alkaloids e.g., akuammicine (II) and strychnine, or by a series d rearrangements generate the Aspidosperma and Ibore alkaloids. Such a model was demonstrated in the laboratory and detailed suggestions made implicating stemmadenine (111) as a key intermediate for this latter process.³ We now describe the isolation and properties of a new alkaloid, preakuammicine, for which the long sought structure IV is proposed and which, from position in the Vinca sequence, meets at least one of the criteria as an intermediate. Furthermore the chemistry of IV provides in vitro analogy for its presumptive role as a true biointermediate.

Careful separation by repeated the of the alkaloid fraction of 42-48-hr-old seedlings of Vinca rosea forded material with homogeneous tlc behavior (0.41 in 20% CHCl₃-MeOH; silica gel G). The amor phous alkaloid preakuammicine, C₂₁H₂₄N₂O₃ (IV) obtained in this way was characterized as an indolening $\lambda_{\text{max}}^{\text{EtOH}}$ 262 nm (ϵ 6000), changing on storage at 20°. λ_{max} 292, 325 nm (chromophore II); nmr (CDCl₃) τ 22 3.0 m (Ar-H); 4.7 q (CH₃CH==); 6.05 s (CH₂OH); 6.1 s (CO_2CH_3); 8.45 d ($CH_3CH=$); mass spectrum (relation)

- (1) A. A. Qureshi and A. I. Scott, Chem. Commun., 945 (1968).
- (2) A. A. Qureshi and A. I. Scott, ibid., 947 (1968).
- (3) A. I. Scott, Chimia, 22, 310 (1968)
- (4) A. I. Scott, 2nd Symposium on Natural Products, Mona, Jamaic Jan 1968.
 - (5) A. A. Oureshi and A. I. Scott, Chem. Commun., 948 (1968).
- (6) E. Wenkert and B. Wickberg, J. Am. Chem. Soc. 87, 1580 (1969 cf. A. R. Battersby, Pure Appl. Chem., 14, 117 (1967); Chimia, 22, 31 (1968).

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tive intensity) M⁺ 352.1757 (2) (calcd for $C_{21}H_{24}N_2O_3$: 352.1787), 322.1683 (100) (calcd for $C_{20}H_{22}N_2O_2$: 322,1682), 263,1560 (25) (calcd for $C_{18}H_{19}N_2$: 263,1549). The mass and infrared spectra of IV were very similar to those reported for the isomeric precondylocarpine⁷ (Y). In particular the mass spectra of both IV and V show base peaks corresponding to the loss of the C_{17} **ca**rbon as formaldehyde to give the $C_{20}H_{22}N_2O_2^+$ species associated with akuammicine (II) and condylocarpine (VI). Treatment of preakuammicine with base (Na-OMe in MeOH, 60°, 15 min) afforded an almost quantitative yield of akuammicine (II) identified by uv, mass spectral, tlc, and ord comparison. This deformylation reaction which finds analogy in the conversion of V to condylocarpine (VI)⁷ thus secures the stereochemistry of IV at positions 3 and 15 as well as simulating the " $C_{10} \rightarrow C_9$ " stage of the biogenesis of the Strychnos alkaloids. In a second model experiment sodium borohydride reduction of IV afforded a separable mixture of akuammicine (II) and stemmadenine (III). The formation of the latter authenticated intermediate of Aspidosperma biosynthesis³ can be rationalized by the well-known⁸ collapse of the

(7) A. Walser and C. Djerassi, *Helv. Chim. Acta*, 48, 391 (1965). We thank Dr. Carl Djerassi for kindly providing us with detailed spectral **data** tor V. system to generate the secoimmonium salt VII. Hydride reduction then affords III which contains the complete carbon skeleton of IV and two of the original stereochemical centers. This experiment provides a satisfactory analogy for the biochemical reduction of IV to III and also for the biochemical interconversion of

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stereochemical centers. This experiment provides a satisfactory analogy for the biochemical reduction of IV to III and also for the biochemical interconversion of IV and V via VII, thus leading to a predicted connection between the *Strychnos* and *Condylocarpine* series. Apart from allocation of the stereochemistry at C₁₆ (which is also unknown in stemmadenine), structure IV follows for preakuammicine. The lability of solutions of IV indicates that this alkaloid has so far defied isolation because of its ready transformation to akuammicine at extremes of pH. From its reactivity and sequence position there seems little doubt that IV and/or the corresponding aldehyde (IV, CH₂OH = CHO)⁹

(9) Preliminary experiments indicate that this aldehyde is also present in immature V. rosea and that IV occurs in the mature plant.

Communications to the Editor

⁽⁸⁾ For a review with many examples see B. Gilbert, Alkaloids, 8, 335 (1965).

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will play a key role in the intermediary metabolism of *V. rosea* and other species.

Finally the mechanism of biogenesis of IV from geissoschizine (1) must be considered. Wenkert⁶ has suggested that coupling of the radical VIII derived from geissoschizine (1) leads to the well-known *akuamma* skeleton IX, rearrangement of IX to IV completing the process. We have previously provided analogy² for a protonation-rearrangement mechanism which with subsequent dehydrogenation connects I and IV. We wish to suggest a third possibility which on the basis of recent model experiments¹⁰ seems to offer an equally attractive rationale and which involves initial oxidation of I to the β -hydroxyindolenine X followed

(10) Unpublished work by Dr. C. R. Bennett,

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by formation of the oxindole XI (R = H) which as the imino ether XI (R = alkyl or phosphate) is well placed to undergo condensation¹¹ to IV.

Experimental test of these possibilities is in progress.

Acknowledgment. Generous support of this work by National Institutes of Health Grant CA-11095 is acknowledged. We thank Dr. W. J. McMurray for the computerized high-resolution mass spectral data.

(11) Cf. T. Oishi, M. Nagai, and Y. Ban, Tetrahedron Letters, 491 (1968).

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